

## 8. New Pterocarpinoids from *Dolichos marginata* ssp. *erecta*

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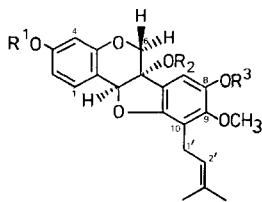
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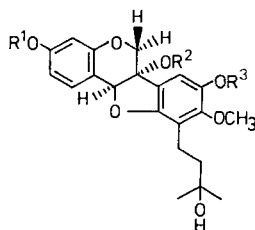
Four new pterocarpinoids, sphenostylin A, B, C, and D (1–4), have been isolated from the CHCl<sub>3</sub> extract of the root bark of *Dolichos marginata* ssp. *erecta* (Leguminosae) by preparative liquid chromatography. The structures have been established by spectroscopic methods (UV, <sup>1</sup>H-NMR, <sup>13</sup>C-NMR, EI-MS, DCI-MS, CD) and chemical transformations. The isolated compounds showed weak antifungal activity against *Cladosporium cucumerinum*.

**Introduction.** – In the course of our systematic screening of African medicinal plants for biological activities, root extracts of *Dolichos marginata* ssp. *erecta* E. MEY. (BAK.) VERDC. (Leguminosae) (syn. *Sphenostylis erecta* E. MEY.) inhibited growth of spores of the fungus *Cladosporium cucumerinum*. The genus *Dolichos* has been studied for the occurrence of lectins, amino acids, and proteins, but the only other constituents of note to be reported are an isoflavone diglycoside [1] and two pterocarpan (= 6a, 11a-dihydro-6*H*-benzofuro[3,2-*c*][1]benzopyrans) and dolichins A and B [2] from *D. biflorus* and saponins from *D. falcatus* [3]. The saponins from *D. falcatus* have analgesic [4] and antitumour activities [5]. In the present paper, the isolation of four antifungal pterocarpinoids from the roots of *D. marginata* ssp. *erecta* is described.

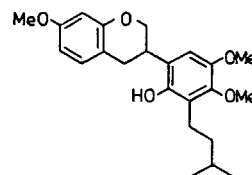
**Results.** – The CHCl<sub>3</sub> extract of *D. marginata* ssp. *erecta* showed by a direct TLC (silica gel, CHCl<sub>3</sub>/MeOH 10:1) bioassay with *Cladosporium cucumerinum* [6] [7] several inhibition zones. The major antifungal components were isolated and characterised.



- 1** R<sup>1</sup> = R<sup>3</sup> = CH<sub>3</sub>, R<sup>2</sup> = H  
**1a** R<sup>1</sup> = R<sup>3</sup> = CH<sub>3</sub>, R<sup>2</sup> = Ac  
**2** R<sup>1</sup> = R<sup>2</sup> = R<sup>3</sup> = H  
**2a** R<sup>1</sup> = R<sup>2</sup> = R<sup>3</sup> = Ac



- 3** R<sup>1</sup> = R<sup>2</sup> = R<sup>3</sup> = H  
**3a** R<sup>1</sup> = R<sup>2</sup> = R<sup>3</sup> = Ac  
**4** R<sup>1</sup> = R<sup>2</sup> = H, R<sup>3</sup> = CH<sub>3</sub>  
**4a** R<sup>1</sup> = R<sup>2</sup> = Ac, R<sup>3</sup> = CH<sub>3</sub>  
**5** R<sup>1</sup> = R<sup>3</sup> = CH<sub>3</sub>, R<sup>2</sup> = H



**6**

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The crude extract was fractionated by medium-pressure liquid chromatography [8] (see *Exper. Part*). The fractions were further purified by low-pressure liquid chromatography to obtain 4 major active compounds, sphenostylins A, B, C, and D (**1–4**). Their molecular formula and structure could be deduced from their MS,  $^1\text{H-NMR}$ , and  $^{13}\text{C-NMR}$  spectra (**1**:  $\text{C}_{23}\text{H}_{26}\text{O}_6$ ; **2**:  $\text{C}_{21}\text{H}_{22}\text{O}_6$ ; **3**:  $\text{C}_{21}\text{H}_{24}\text{O}_7$ ; **4**:  $\text{C}_{22}\text{H}_{26}\text{O}_7$ ) and from chemical transformations.

The 4  $^1\text{H-NMR}$  spectra showed a typical pattern assigned to the heterocycle protons of 6a-hydroxypterocarpan. This included an *AB* pattern ( $q'$ , 2 H–C(6)) at *ca.* 4 ppm and a low-field *s* (H–C(11a)) at *ca.* 5.2 ppm (see *Table*), as observed for compounds of this class [9]. This was confirmed by easy conversion into their pterocarpene derivatives, a feature characteristic of 6a-hydroxypterocarpan [10]. The reaction was achieved by adding 1 drop of conc. HCl to a solution of pterocarpan in a UV cell. We observed the following bathochromic shifts: 285 to 340 nm for **1**, 284 to 335 nm for **2**, 285 to 335 for **3**, and 286 to 335 nm for **4**. The sphenostylins **1–4** exhibited furthermore an aromatic *ABC* pattern and an aromatic *s*. The presence of 3 MeO groups and an isopentenyl substituent for **1** and 2 MeO groups and an isopentenyl function for **2** is readily apparent (see *Table*). Compounds **3** and **4** have 1 and 2 MeO substituents, respectively. They show, in addition, the presence of a  $(\text{CH}_3)_2\text{C}$  group (1.1 ppm for **3**; 1.2 ppm for **4**), and *m*'s at 1.45 and 2.45 ppm for **3** and 1.6 and 2.65 ppm for **4**, characteristic signals for 3-hydroxyisopentenyl groups.

Treatment of **1–4** with  $\text{CH}_2\text{N}_2$  gave the following results: Compound **1** was unaffected, and **2** yielded a substance identical to **1** ( $^1\text{H-NMR}$ , UV, MS); **3** and **4** yielded the same compound **5** ( $^1\text{H-NMR}$ , UV, MS). The permethylated compounds **1** and **5** exhibited the same aromatic *ABC* pattern (chemical shifts and coupling constants; see *Table*). The values observed are comparable with those of 3-substituted pterocarpan [9]. Thus, substances **1–4** must have the same substitution on ring A. The monosubstitution of ring A was confirmed by hydrogenolysis (10% Pd/C, AcOEt) of **1**. The obtained isoflavan **6** was analyzed by MS. It displayed a typical isoflavan fragmentation pattern [11] with major peaks at  $m/z$  250 (46%),  $m/z$  249 (52%), and  $m/z$  137 (28%).

The positions of the ring-D substituents were determined by acetylation of **1–4** (see *Exper. Part*). The acetates **1a–4a** were unstable (formation of the corresponding pterocarpenes) upon separation on silica gel, but could be purified by rapid *Lobar* chromatography (silica gel) for **2a** and **3a** and by HPLC (*RP-18*) for **4a** and **1a**. In each case, the phenolic functions and the OH group at C(6a) were acetylated. For **3** and **4**, the tertiary OH function in the side chain remained unaffected.

The  $^1\text{H-NMR}$  chemical shifts of **1a–4a** permitted to deduce the positions of the different substituents. For the aromatic ring A, a MeO group is located at C(3) for **1** and an OH function at C(3) for **2–4** (see *Table*; chemical-shift displacement for H–C(1), H–C(2), and H–C(4) in **2a–4a**). In addition, we observed an important downfield shift of the proton at ring D. In **1a** and **4a**, the shift ( $\Delta\delta = 0.3$  ppm) is only due to the contribution of the AcO group at C(6a) (no phenolic function on ring D for **1a** and **4a**). For compounds **2a** and **3a**, the shift ( $\Delta\delta = 0.45$  ppm) is due to the presence of 2 AcO moieties, one at C(6a) ( $\Delta\delta = 0.3$  ppm) and one at ring D ( $\Delta\delta = 0.15$  ppm). From these values, it can be concluded that the aromatic proton at ring D is next to C(6a) and thus at C(7) for **1–4**.

Finally, from biosynthetic considerations [6] and comparison of chemical shifts with known pterocarpan [9] [12] [13], the isopentenyl and hydroxyisopentenyl substituents can be located at C(10). Hence, C(9) of ring D is occupied by the remaining MeO group in all 4 compounds.

The sphenostylins A, B, C, and D **1–4** are new 6a-hydroxypterocarpan. Measurement of their CD spectra [9] showed for each compound a strong negative *Cotton* effect

Table. <sup>1</sup>H-NMR Spectra (200 MHz)

	H-C(1)	H-C(2)	H-C(4)	H-C(7)	H-C(8)
Cristacarpin [9]	7.33 ( <i>d</i> , <i>J</i> = 8.5)	6.56 (' <i>q</i> ', <i>J</i> = 8.5, 2.3)	6.31 ( <i>d</i> , <i>J</i> = 2.3)	7.18 ( <i>d</i> , <i>J</i> = 8)	6.54 ( <i>d</i> , <i>J</i> = 8)
<b>1</b> (CDCl <sub>3</sub> )	7.42 ( <i>d</i> , <i>J</i> = 8)	6.65 (' <i>q</i> ', <i>J</i> = 8, 2)	6.45 ( <i>d</i> , <i>J</i> = 2)	6.79 ( <i>s</i> )	–
<b>2</b> (CDCl <sub>3</sub> )	7.32 ( <i>d</i> , <i>J</i> = 7)	6.55 (' <i>q</i> ', <i>J</i> = 7, 2)	6.37 ( <i>d</i> , <i>J</i> = 2)	6.82 ( <i>s</i> )	–
<b>1a</b> (CDCl <sub>3</sub> )	7.40 ( <i>d</i> , <i>J</i> = 8)	6.65 (' <i>q</i> ', <i>J</i> = 8, 2)	6.42 ( <i>d</i> , <i>J</i> = 2)	7.12 ( <i>s</i> )	–
<b>2a</b> (CDCl <sub>3</sub> )	7.51 ( <i>d</i> , <i>J</i> = 8)	6.82 (' <i>q</i> ', <i>J</i> = 8, 2)	6.68 ( <i>d</i> , <i>J</i> = 2)	7.26 ( <i>s</i> )	–
<b>3</b> ((D <sub>6</sub> )DMSO)	7.20 ( <i>d</i> , <i>J</i> = 8)	6.40 (' <i>q</i> ', <i>J</i> = 8, 2)	6.20 ( <i>d</i> , <i>J</i> = 2)	6.67 ( <i>s</i> )	–
<b>4</b> (CD <sub>3</sub> OD)	7.28 ( <i>d</i> , <i>J</i> = 8)	6.48 (' <i>q</i> ', <i>J</i> = 8, 2)	6.27 ( <i>d</i> , <i>J</i> = 2)	6.88 ( <i>s</i> )	–
<b>5</b> (CDCl <sub>3</sub> )	7.41 ( <i>d</i> , <i>J</i> = 8)	6.65 (' <i>q</i> ', <i>J</i> = 8, 2)	6.45 ( <i>d</i> , <i>J</i> = 2)	6.80 ( <i>s</i> )	–
<b>3a</b> (CDCl <sub>3</sub> )	7.51 ( <i>d</i> , <i>J</i> = 9)	6.81 (' <i>q</i> ', <i>J</i> = 9, 2)	6.67 ( <i>d</i> , <i>J</i> = 2)	7.25 ( <i>s</i> )	–
<b>4a</b> (CDCl <sub>3</sub> )	7.51 ( <i>d</i> , <i>J</i> = 8)	6.82 (' <i>q</i> ', <i>J</i> = 8, 2)	6.66 ( <i>d</i> , <i>J</i> = 2)	7.10 ( <i>s</i> )	–

<sup>a</sup>) Chemical shifts are given in ppm and coupling constants *J* in Hz.

(see *Exper. Part*), thus indicating the (6*aS*,11*aS*)-configuration. In a TLC bioassay with the fungus *Cladosporium cucumerinum*, the pure compounds inhibited spore growth at a minimum concentration of 6.25 μg for **1**, 10 μg for **2**, 20 μg for **4**, and 50 μg for **3**.

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### Experimental Part

*General.* Acetylations were carried out by stirring the compounds with Ac<sub>2</sub>O in pyridine for 24 h at r.t. Medium-pressure liquid chromatography (MPLC): Büchi B-681 system. Low-pressure liquid chromatography (LPLC): Lobar silica-gel column (Merck). HPLC: Perkin Elmer Series 3B, column RP-18 Knauer semi-prep. TLC: silica-gel precoated Al sheets (Merck), CHCl<sub>3</sub>/MeOH 9:1, visualisation with Godin reagent [14]. UV: Perkin Elmer Lambda 3. CD (λ(Θ) in nm): Jasco J-500 C. <sup>1</sup>H- and <sup>13</sup>C-NMR spectra; Bruker WP-200 (200 and 50.29 MHz) in CDCl<sub>3</sub>, CD<sub>3</sub>OD, or (D<sub>6</sub>)DMSO using TMS as an internal standard. MS: Nermag R-3010 spectrometer.

*Plant Material.* *D. marginata* ssp. *erecta* was collected at Zomba, Malawi. A voucher specimen of the plant material is retained at the Herbarium, Chancellor College, University of Malawi, Zomba.

*Extraction and Isolation.* The powdered root bark of *D. marginata* (152 g) was extracted at r.t. with petroleum ether (625 mg), followed by CHCl<sub>3</sub> (11 g). A part of the crude extract (3.5 g) was separated by MPLC on a 36 mm × 92 cm column (silica gel Merck 9385) with CHCl<sub>3</sub>/MeOH 10:0.8. Five fractions were collected. *Fraction 1* (140 mg) was further purified on a Lobar silica-gel column with petroleum ether/AcOEt 5:5 affording **1** (62 mg). *Fraction 4* (580 mg) was purified by MPLC on a 36 mm × 92 cm column (silica gel), with petroleum ether/AcOEt 3:7: **2** (43 mg) and **3** (35 mg). *Fraction 5* was purified on a Lobar silica-gel column with petroleum ether/AcOEt 2:8: pure **4** (27 mg).

*Sphenostylin A* (= (6*aS*,11*aS*)-6*a*,11*a*-Dihydro-3,8,9-trimethoxy-10-(3-methyl-2-butenyl)-6*H*-benzofuro-[3,2-*c*] [1]benzopyran-6*a*-ol; **1**). Transparent oil. TLC (SiO<sub>2</sub>): R<sub>f</sub> 0.60. UV (CHCl<sub>3</sub>): 243, 285, 298. CD (CHCl<sub>3</sub>): 244 (–35000), 277 (–5700), 286 (0), 290 (+3400), 297 (0), 305 (–1700). <sup>1</sup>H-NMR: Table. <sup>13</sup>C-NMR (CDCl<sub>3</sub>): 17.96

*Sphenostylins A–D (1–4) and of their Derivatives<sup>a)</sup>*

H–C(6)	H–C(11a)	2 H–C(1')	1 or 2 H–C(2')	2 CH <sub>3</sub> –C(3')	MeO	AcOAr	AcO–C(6a)
.06, 4.16 <i>AB, J = 12</i>	5.27 ( <i>s</i> )	3.25 ( <i>d, J = 7</i> )	5.16 ( <i>t, J = 7</i> )	1.65 ( <i>s</i> ) 1.76 ( <i>s</i> )	3.83 ( <i>s</i> )	–	–
.05, 4.22 <i>AB, J = 11</i>	5.24 ( <i>s</i> )	3.25 ( <i>d, J = 7</i> )	5.17 ( <i>t, J = 7</i> )	1.63 ( <i>s</i> ) 1.73 ( <i>s</i> )	3.77 ( <i>s, 6H</i> ) 3.82 ( <i>s</i> )	–	–
98, 4.18 <i>AB, J = 12</i>	5.20 ( <i>s</i> )	3.27 ( <i>d, J = 7</i> )	5.17 ( <i>t, J = 7</i> )	1.65 ( <i>s</i> ) 1.73 ( <i>s</i> )	3.75 ( <i>s</i> )	–	–
51, 4.69 <i>AB, J = 12</i>	5.58 ( <i>s</i> )	3.25 ( <i>d, J = 7</i> )	5.17 ( <i>t, J = 7</i> )	1.63 ( <i>s</i> ) 1.73 ( <i>s</i> )	3.77 ( <i>s</i> ) 3.78 ( <i>s</i> ) 3.83 ( <i>s</i> )	–	2.06 ( <i>s</i> )
35, 4.78 <i>AB, J = 12</i>	5.59 ( <i>s</i> )	3.27 ( <i>d, J = 7</i> )	5.14 ( <i>t, J = 7</i> )	1.65 ( <i>s</i> ) 1.73 ( <i>s</i> )	3.75 ( <i>s</i> )	2.30 ( <i>s</i> ) 2.29 ( <i>s</i> )	2.06 ( <i>s</i> )
80, 4.00 <i>AB, J = 11</i>	5.05 ( <i>s</i> )	2.45 ( <i>m</i> )	1.45 ( <i>m</i> )	1.10 ( <i>s</i> )	3.65 ( <i>s</i> )	–	–
92, 4.10 <i>AB, J = 12</i>	5.15 ( <i>s</i> )	2.60 ( <i>m</i> )	1.60 ( <i>m</i> )	1.19 ( <i>s</i> ) 1.20 ( <i>s</i> )	3.75 ( <i>s</i> ) 3.80 ( <i>s</i> )	–	–
02, 4.22 <i>AB, J = 11</i>	5.23 ( <i>s</i> )	2.65 ( <i>m</i> )	1.67 ( <i>m</i> )	1.23 ( <i>s</i> )	3.77 ( <i>s</i> ) 3.80 ( <i>s</i> ) 3.83 ( <i>s</i> )	–	–
35, 4.74 <i>AB, J = 12</i>	5.60 ( <i>s</i> )	2.62 ( <i>t, J = 8</i> )	1.68 ( <i>m</i> )	1.23 ( <i>s</i> ) 1.24 ( <i>s</i> )	3.77 ( <i>s</i> )	2.28 ( <i>s</i> ) 2.29 ( <i>s</i> )	2.05 ( <i>s</i> )
49, 4.71 <i>AB, J = 12</i>	5.50 ( <i>s</i> )	2.65 ( <i>t, J = 7</i> )	1.68 ( <i>m</i> )	1.22 ( <i>s</i> )	3.82 ( <i>s</i> ) 3.83 ( <i>s</i> )	2.28 ( <i>s</i> )	2.05 ( <i>s</i> )

(CH<sub>3</sub>–C(3')); 23.66 (C(1')); 25.86 (CH<sub>3</sub>–C(3')); 55.64 (CH<sub>3</sub>O–C(3)); 57.12 (CH<sub>3</sub>O–C(8)); 60.97 (CH<sub>3</sub>O–C(9)); 70.00 (C(6)); 78.08 (C(6a)); 84.4 (C(11a)); 102.08 (C(7)); 105.83 (C(4)); 110.00 (C(2)); 113.28 (C(11b)); 120.30 (C(6b)); 121.92 (C(1)); 122.59 (C(2')); 131.95 (C(10)); 132.35 (C(3')); 148.70 (C(8)); 150.36 (C(9)); 152.76 (C(10a)); 156.14 (C(4a)); 161.53 (C(3)). EI-MS: 398 (100, *M*<sup>+</sup>), 380 (18), 370 (20), 342 (14), 314 (20), 299 (18), 177 (27).

Acetate **1a** was purified by HPLC on *RP-18* with MeOH/H<sub>2</sub>O 95%→100% MeOH in 20 min. <sup>1</sup>H-NMR: *Table*. DCI-MS (NH<sub>3</sub>): 440 (*M*<sup>+</sup>).

6-(3,4-Dihydro-7-methoxy-2H-1-benzopyran-3-yl)-3,4-dimethoxy-2-(3-methylbutyl)phenol (**6**). For 24 h, 1.5 mg of **1** was hydrogenated over 10% Pd/C in AcOEt. After filtration, the mixture was purified on silica gel with petroleum ether/AcOEt 9:1. The main component **6** was analyzed by MS. EI-MS: 386 (100, *M*<sup>+</sup>), 249 (52), 250 (46), 137 (28).

*Sphenostylin B* (= (6*aS*,11*aS*)-6*a*,11*a*-Dihydro-9-methoxy-10-(3-methyl-2-butenyl)-6H-benzofuro[3,2-*c*]/[1]benzopyran-3,6*a*,8-triol; **2**). Oil. TLC (SiO<sub>2</sub>): *R*<sub>f</sub> 0.45. UV (CHCl<sub>3</sub>): 245, 284, 298. CD (CHCl<sub>3</sub>): 246 (–15700), 279 (–3800), 285 (O), 289 (+2400), 294 (O), 303 (–1400). <sup>1</sup>H-NMR: *Table*. <sup>13</sup>C-NMR (CDCl<sub>3</sub>): 17.85 (CH<sub>3</sub>–C(3')); 23.63 (C(1')); 25.66 (CH<sub>3</sub>–C(3')); 61.49 (CH<sub>3</sub>O–C(9)); 69.59 (C(6)); 77.73 (C(6a)); 83.98 (C(11a)); 103.80 (C(7)); 107.45 (C(4)); 110.41 (C(2)); 113.21 (C(11b)); 118.99 (C(6b)); 121.94 (C(1)); 123.21 (C(2')); 132.26 (C(10)); 132.50 (C(3')); 144.12 (C(8)); 147.36 (C(9)); 151.88 (C(10a)); 155.95 (C(4a)); 157.26 (C(3)). DCI-MS (NH<sub>3</sub>): 370 (*M*<sup>+</sup>).

Triacetate **2a** was purified by chromatography on silica gel with petroleum ether/AcOEt 7:3. <sup>1</sup>H-NMR: *Table*. DCI-MS (NH<sub>3</sub>): 496 (*M*<sup>+</sup>).

*Sphenostylin C* (= (6*aS*,11*aS*)-6*a*,11*a*-Dihydro-10-(3-hydroxy-3-methylbutyl)-9-methoxy-6H-benzofuro[3,2-*c*]/[1]benzopyran-3,6*a*,8-triol; **3**). TLC (SiO<sub>2</sub>): *R*<sub>f</sub> 0.37. UV (MeOH): 230, 280 (sh), 285, 298. CD (MeOH): 249 (–20500), 286 (–2150), 290 (O), 296 (+2400), 305 (O), 313 (–1700). <sup>1</sup>H-NMR ((D<sub>6</sub>)DMSO): *Table*. <sup>13</sup>C-NMR ((D<sub>6</sub>)DMSO): 19.08 (2 CH<sub>3</sub>–C(3')); 28.94 (C(2')); 43.41 (C(1')); 60.04 (CH<sub>3</sub>O–C(9)); 68.90 (C(6)); 69.40 (C(3')); 75.85 (C(6a)); 83.22 (C(11a)); 102.70 (C(7)); 108.65 (C(4)); 109.86 (C(2)); 111.98 (C(11b)); 119.18 (C(6b)); 123.61 (C(1)); 132.02 (C(10)); 144.24 (C(8)); 147.18 (C(9)); 150.34 (C(10a)); 155.60 (C(11a)); 158.62 (C(3)). DCI-MS (NH<sub>3</sub>): 388 (*M*<sup>+</sup>).

Triacetate **3a** was purified on silica gel with petroleum ether/AcOEt 5:5. <sup>1</sup>H-NMR: *Table*. DCI-MS (NH<sub>3</sub>): 514 (*M*<sup>+</sup>).

(6*aS*,11*aS*)-6*a*,11*a*-Dihydro-10-(3-hydroxy-3-methylbutyl)-3,8,9-trimethoxy-6H-benzofuro[3,2-*c*][1]benzopyran-6*a*-ol **5**. Compound **3** or **4** was treated in MeOH with an excess of ethereal diazomethane: **5**. <sup>1</sup>H-NMR: Table. UV (CHCl<sub>3</sub>): 242, 280 (sh), 285, 297. DCI-MS (NH<sub>3</sub>): 416 (*M*<sup>+</sup>).

Sphenostylin *D* (= (6*aS*,11*aS*)-6*a*,11*a*-Dihydro-10-(3-hydroxy-3-methylbutyl)-8,9-dimethoxy-6H-benzofuro[3,2-*c*][1]benzopyran-3,6*a*-diol; **4**). TLC (SiO<sub>2</sub>): *R*<sub>f</sub> 0.31. UV (MeOH): 230, 280 (sh), 286, 295 (sh). CD (MeOH): 248 (–27300), 283 (–2000), 292 (O), 297 (+1600), 305 (O), 310 (–800). <sup>1</sup>H-NMR: Table. <sup>13</sup>C-NMR (CD<sub>3</sub>OD): 20.37 (2CH<sub>3</sub>–C(3′)); 29.04 (C(2′)); 44.83 (C(1′)); 57.51 (CH<sub>3</sub>O–C(8)); 61.53 (CH<sub>3</sub>O–C(9)); 70.84 (C(6)); 71.68 (C(3′)); 78.21 (C(6*a*)); 85.57 (C(11*a*)); 104.23 (C(7)); 107.45 (C(4)); 111.29 (C(2)); 113.58 (C(11*b*)); 121.55 (C(6*b*)); 124.42 (C(1)); 133.36 (C(10)); 149.49 (C(8)); 150.78 (C(9)); 153.68 (C(10*a*)); 157.49 (C(11*a*)); 160.21 (C(3)). DCI-MS (NH<sub>3</sub>): 402 (*M*<sup>+</sup>).

Diacetate **4a** was purified by HPLC on *RP-18* with MeOH/H<sub>2</sub>O 90% → 100% MeOH in 20 min. <sup>1</sup>H-NMR: Table. DCI-MS (NH<sub>3</sub>): 486 (*M*<sup>+</sup>).

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